

REplication Jumping Duration PCR Amplification,Closing and Sanger Sequencing of A Fragment of the Temptation Gene in Black-spotted frog1

Xindi Cui

Inner Mongol University of Technology, Shijiazhuang, Hebei,Province

Abstract: During in vitro replication of DNA by DNA Polymerase,The diverse palindromic StructuresOn templates of DNA can lead to replication jumping and cause fragments to be missing from daughter strands.We recorded replication jumping in which a 564 bp fragment was missing from daughter DNA.The original template was a 1126 bp fragment of the introduction of the temporary gene of black-spotted frog(*Pelophylax nigromaculatus*).The replication jump was recorded after amplification of this template using polymerase chainREaction,Closing Using Vector,And sequencing using the Sanger Method.The replication jump was elevated to rise from this series of events:

Keywords: Reloading DNA polymerase; Replication; vitro processes; fragment; replication jump

Polymerase Chain Reaction(Polymerase ChainREactionPCR)Is refers to inDNAPolymerase of catalytic under to mother chainDNAFor template to specific primers for extension starting point by degeneration,Annealing,Extension and steps in vitro copy the and mother chain TemplateDNAComplementary of sub-chain DNAOf process is a can fast,Specifically in vitro amplification objectiveDNAOf technology is biological research in application the most widely [2-4] Laboratory Technology one of PCRProduct often need to After cloning after the sequencing or other Analysis.Cloning an arcane willPCRProduct and specific carrierDNAMolecular connection formation recombinantDNAMolecular into competent cells in by cells and Carrier of emerging suchT-Carrier and in very big degree on the simplified.PCRProduct of cloning process at the same time also improve the cloning Effect [8-10] Rate But,PCRNot always completely faithful of In addition to primers and template of non-specific binding was to produce non-specific opposite sex amplification product outside because template of superior structure,Polymerase of some properties such as also often will mediated-DNAOf recombinant(Recombination),Copy sliding(Slippage)And

Recombinant isDNAPolymerase in In accordance with template synthesis new chain of when because the other a and template sequence height similar of fragment and template in space on the close, polymerase from original of template chain on the move to the fragment on the continue to synthesis New Chain Final makePCRProduct become two different template chain[12]

The recombinant product.Copy sliding in microsatellite and short tandem repeat sequence of amplification in very common,DNAPolymerase in synthesis new chain more. A or a few repeat unit causedPCRProduct than template chain shorten[13].

Repeat unit integer times Jump amplification is when templateDNATheir own because the original sequence formation more stability of ring tongue local secondary structure New Chain Synthesis whenDNAPolymerase move to the

Copyright © 2019 .

This is an open-access article distributed under the terms of the Creative Commons Attribution Unported License

(<http://creativecommons.org/licenses/by-nc/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

regional after it can span this ring tongue area continue to down synthesis new chain to make PCR Product is template lost the

Amplification people 18 S rDNA Gene when found the middle missing 54 BPOf abnormal.

Is big fragment of jump more difficult to happen in experimental also very rare. Because so, PCROf jump problem often be ignored caused error results.

PCR Product of cloning is use bacteria in DNA Polymerase and copy system will target DNA Fragment the in body copy. Because bacteria Copy System of faithful of much higher than in general Taq DNA Polymerase (No 3'-5' Enzyme Activity) Molecular cloning using plasmid replication is generally considered to be very accurate. However, in special cases, such as when the target molecule contains some special palindrome structure, the bacterial plasmid.

Jumps or slides during replicatio.

We're studying the black-spotted frog (*Pelophylax nigromaculatus*) Antimicrobial Peptides Temporin When it comes to gene structure PCR Amplification and amplification products T-A Jumps occur simultaneously during cloning and cause up

BP The fragment is missing and found with a segment 6 BP About the repeat unit. We reported the results of this experiment to provide a reference for more research..

1. Materials and Methods

1.1 Sample

Chuan, Xi'an, Taiyuan, etc. 20 Adult black spotted lateral pleats collected in different regions, and each region 5. Only, the hind leg muscle samples were taken immediately after double-damaged pulp was executed in the wild and preserved in liquid nitrogen, and then transferred -80°C in the refrigerator Save.

1.2 General DNA Extraction

20 mg Add muscle tissue after being cut 25 µL protein-ase K (10 mg/ml), 35 µL SDS (10%), 295 µL the Buffer (PH = 8), Fully mix and then centrifuge, 56°C Water bath digestion

H. Digestive Juice Axy prep Genome DNA Extraction kit (Axy prep, Hangzhou) Total Extraction DNA, Operation in accordance with the instructions.

As shown in figure 1 PCR Product by agarose electrophoresis found big about 73% Of individual in 1200 BP And 550 BP The at the same time appear clear of two, 18% Individual short of strip signal weaker, 9% The individual only 1200 BPOf Strip. Will two the bands respectively recovery after and sequencing results display is long of bands 1126 BP Short of bands 562 BP After is in the former 198 bp The start missing 564 bp Made. ference for two different of allele gene.

But in the long fragment Sanger Sequencing when found a kind of abnormal phenomenon template actual length 1126 BP But sequencing results in sometimes will only get 562 BPOf Short Sequence. The short sequence is long fragment in BP The start missing 564 bp Made. This show that short very may is PCR Cause of a kind of illusion the in amplification reaction in the fragment missing.

With recovery of long fragment as an Template PCR Amplification agarose gel electrophoresis detection PCR Product when found 3A situation A is only get 1126 BP The long fragment A is only get 562 BPOf Short Fur ther more two fragment at the same time (Figure 2) On PCR Product sequencing also prove that the this a little.

Template pollution of possibility we the long fragment purification recovery after. T-A Cloning. Extraction positive cloning of plasmid after with restriction enzyme Pst I And Not I To PCR Product from carrier on cut down again by agarose electrophoresis detection found positive cloning in containing two kind of cloning fragment A is containing and long fragment completely same of cloning Fragment, A kind of IS and short completely same of cloning fragment (figure 3). That is, 1126 BP Long of objective fragment in different colony in become size two of Fragment. Sequencing show that short remain is long fragment 198 bp The missing 564 bp Made. This show that this 1126 BP Long of fragment not only in PCR Amplification process in can happen missing and in *Escherichia coli* of in of plasmid copy process also can

occur missing this a kind of phenomenon is very rare.

Use software REPEAT around 2.10kay1126 BPOf fragment the repeat sequence analysis results display the sequence has the same direction repeat sequence (Direct repeats) 70Of, Reverse repeat sequence (Inverse re-peats) 13Of, Mirror repeat sequence (Mirror repeats) 21 Of and complementary repeat sequence (Complementary repeats) 21 Of (Table1). Will1126 BPAnd526 BPThe sequence of the two fragments was compared and found that the two sequences at the upstream and end of the missing fragment have one6 BPSame-direction repeat sequence 5'-AGACCT-3'(Figure4.)Is where the sequence deletion occurred, so far we speculate DNAA jump occurred while copying to the second repeat sequence (PCRJumping).

2. Discussion

DNAA sequence can be repeated in the same direction, in the same direction[20] Jump or slide. In this study 1126 BPDetected in Fragment Different repetitive sequences (Table1.), But repeated amplification, only in 194~199 BPLocation and first 757~762 BPRepeat sequence that appears on the location 5'-AGACCT-3'A jump occurred at the site, resulting in the deletion of the amplified product 564 bp. Surprisingly, by this pair of repetitions The jump will occur in vitro and in vivo[21.]

CancellTwo replication-hopping models are proposed..One is related to the double strand hairpin structure of the parent..The strand synthesis is closed at the base that forms the hairpin structure, and then through DNAPolymerase Chain replacement activity continued and finally slid over the hairpin Structure. However, the parental double helix model does not seem to be applicable to our replication Jump Events because in our study, a repeat sequence is missing in the sub chain. This indicates that one of the repeat sequences is incorporated into the hairpin structure,

Yes, in our study, the sequence after the first repeat or before the second repeat cannot form a stable hairpin structure with the repeat sequence (Figure5AAnd5b).

The other model is the heterohelix model, which blocks the synthesis of the chain at the first repeat, while DNAPolymerase and sub-chain are detached from the template. In the next cycle, the new chain acts as a complementary chain, matching the second repeat unit., Annealed and became primers, DNAPolymerase rebinds, starting downstream of the second repeat unit to complete the new chain

Because in this case a repeat sequence is missing in the sub chain, which is consistent with our case..The only difference between our results and the model is the weak double chain body at the bottom of the hairpin. It is reported that, DNAPolymerase shedding and chain extension blocking may not be caused by hairpin, but DNA

This structure appears when the copy occurs, which leads to the occurrence of jumps (Figure5C). We were unable to test this prediction further, but we speculated that both high-level structural-induced strand synthesis blocking and Polymerase shedding were associated with repeat sequences, and this happens both in vivo and in vitro..

DNAThe ability of Polymerase to produce jumping is related to its chain replacement activity. When plasmid replication is done in bacteria, DNAPolymerase III (Early replication, sometimes DNAPolymerase I)^[24] When making a new chain extension, because of its chain^[25],

Replacement activity is not high Appropriate advanced knot on encountered Template After shedding, the new chain can be reannealed with another homologous sequence (the second repeat unit in this study, and finish the chain extension and jump. Single clone in this study DNAThis is probably the case when the fragment is cloned again. (Figure5C) In fact PCR, Sanger Sequenced or Molecular Cloned DNA The type of Polymerase Taq DNA Polymerase, DNA Polymerase II, T4 Polymerase, etc.

A high structure that is easy to jump and is also prone to replication jumps when these enzymes are involved in replication.

The replication jump encountered by this research not only happens in Temporalin The two-way amplification of the gene intron also occurs in Sanger In the process of sequencing, it even happened in the Process of plasmid replication in bacteria.. Based on our confirmation experiment, the purified 1126 BP The fragment was used as a template for

reamplification and the positive plasmid was digested with double enzymes, which led to this conclusion; otherwise, jumping may be misunderstood as an independent allele. So, we suggest that when we use Fragment Size for diploid or polyploid genotyping, we should first select the allele with larger fragment

References

1. Li meiyang. DNA In vitro amplification technology---Polymerase Chain Reaction (PCR) J. Shen Journal of Yang Normal University: Natural Science, 2000, 18(4.): 40-44.
2. Guo yulian, PCR Principle and Application of Technology J. Wisdom, 2008(18): 230, 168.
3. Lynch J. PCR Technology: Principles and Applications for DNA Amplification J. Journal of Medical Genetics, 1990, 27(8.): 536.
4. Beroldingen C H V, Blake E T, Higuchi R, et al. Applications of PCR to the analysis of biological evidence M. PCR Technology. London: Palgrave Macmillan, 1989: 209-223.
5. Xu Yishan, He Jingjing, Chen wanjuan, et al. The Principle and Application of cloning technology J. Technology Outlook, 2015(6.): 89-90.
6. Sambrook J, Russell D W. Molecular cloning experiment Guide M. Huang Pei Hall Translation., Beijing: Chemical Industrial Press, 2008.
7. Xue Fen ground Xu Qing Wei Hua and. Only PCR The Molecular Cloning of Methods explore J. Capital Medical University of Journal, 2014 35(6): 785-789.
8. ROBL J M. Application. cloning technology. production. Human polyclonal antibodies. cattle J. Cloning, Stem Cells 2007 9(1): 12-16.
9. Yellow yuan Tao Ying Zhang Wen dew and. A kind of new DNA Molecular cloning methods J, China Science: Life Science, 2011 41(9): 722-729.
10. Lewis I M Peura T Trounson A O. Large-scale applications. Cloning technologies. agriculture: An industry perspective J. REproduction Fertility, development 1998 10(8): 677-682.
11. Luo Rui. Continuous PCR Amplification process in abnormal phenomenon of Mechanism Research D. North Beijing: China Academy of Sciences Plant Institute, 2006.
12. Saiki R K Gelfand D H Stoffel S E et al. Primer-directed enzymatic amplification DNA. a thermostable DNA Polymerase J. Science 1988 239(4839): 487-491.
13. Hartenstine M J Goodman M F Petruska J. Base stacking and even/odd behavior of hairpin loops in DNA triple repeat slippage and expansion with DNA Polymerase J. The Journal of Biological Chemistry, 2000, 275(24): 18382-18390.