

# Polyethyleneimine Enhances *in situ* Gene Delivery, Expression. Skeletal Muscles. BALB/c mice. Pluronic-mediated System

Jingjing Hu

University of Engineering, Shijiazhuang, Hebei Province, 050100;

**Abstract:** Objective. correctly introduce cationic polyethyleneimine(PEI). Pluronic L64-mediated gene de-livery system. mice skeletal muscle. enhance. efficiency. gene expression. methods. study Different PEI samples. transferred. mice tibialis anterior muscles via intramuscular injection. Saline Plasmid DNA(PDNA) And this may contribute to the process of gene de-livery in skeleton muscle-Based System, Although pDNA was not fully compressed in L/P/D-0.5 group. Conclusion The L/P/D-0.5 system can safely and effectively improved the expression of existing genes in skeleton muscle.

**Keywords:** BALB/c mice; Skeleton Muscle gene delivery; Polyethyleneimine; Pluronic

Skeletal Muscle Distribution, The gene encoding functional protein is transferred to skeletal muscle cells by intramuscular injection. "Factory" In the patient "Production" Therapeutic protein, achieving individual, Precision therapy is a significant and promising treatment strategy (Lu *et al.*, 2003a). How to safely and efficiently transfer foreign genes into muscle cells is the key to the success of this strategy. Although the plasmid DNA (PDNA) can be transferred to muscle cells for expression, but due to direct injection PDNA easily degraded, Transfection efficiency is not ideal due to the low efficiency. Therefore, people have been devoted to developing various gene transfer strategies to improve the efficiency of Gene Transfer and Expression in muscle, such as e-transfer method, Ultrasound and carrier mediated methods (MIR *et al.* 1998; Gu, Rin 2000; Lauritzen *et al.* 2002; Lu *et al.* 2003b; Wells 2004; Lee *et al.* 2012). This paper explore the cationic polymer---Polyethylenimine (Polyethyleneimine PEI) As an gene carrier in skeletal muscle *in situ* Gene Transfer System in application of possibility use PEI/pDNA Complex and Pluronic L64 Combined with construction the Security efficient of Gene Transfer System (L/P/D) Which, LS Pluronic L64 P Representative PEI D Representative PDNA. By the Different Nitrogen and Phosphorus (N/P) Of L/P/D System mediated-of gene expression efficiency and in biological safety evaluation reveal N/P And muscle in gene transfection efficiency of relationship and established PEI In skeletal muscle in application of effective programme. The programme for screening and Development efficient, For use with clinical treatment of skeletal muscle *in situ* gene transfer system provide the appropriate of principle and Strategy.

## 1. Material and Methods

### 1.1 Experimental Animal

5~7/Weeks age body quality Natural 20~25g Of BALB/c Male Mice some purchased in Chengdu da shuo biological science and technology limited the company [Experimental Animal Production License number: Syxk (Sichuan) 2015-030]. Animal Experiment were in Sichuan University ethics committee recognized under the in

Copyright © 2019 .

This is an open-access article distributed under the terms of the Creative Commons Attribution Unported License

(<http://creativecommons.org/licenses/by-nc/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

accordance with the relevant regulations and system [experimental animal "with license number: Syxk (Sichuan) 2013-017]. Will mice in 21, Humidity 45%~65% Of environment in random cage feeding. Every group 6 Only all mice were injection bilateral leg Tibialis Anterior Muscle.

## 1.2 Reagent and Instrument

A of polyethylenimine 25 kDa, Pluronic L64 (Sigma-Aldrich American);  $\beta$ -Galactosidase (PCMV-LacZ) In situ Staining Kit (PIK Wanjia, China); E.Z.N.A.<sup>TM</sup> Plasmid small extraction kit, Purelink<sup>TM</sup> High purity plasmid a lot extraction kit

$\beta$ -Galactosidase, Fluorescein enzyme (PCMV-Luc) And far-infrared fluorescence protein (PCMV-E2) Of 3A plasmid by this laboratory Liu yili Dr. provide. Chemidoc<sup>TM</sup> XRS + Gel imaging system (Bio-Rad American); Varioskan Flash Multi-function microplate reader (Thermo American); Nanos ZEN 1600 Style nano-particle size and potential analyzer Malvern British); NanoDrop 2000 (Thermo American); MFP-3D-BIO<sup>TM</sup> Atomic Force Microscope (Bruker American); - Vivo Imaging System (CRI American).

## 1.3 Experimental Methods

### 1.3.1 Plasmid of preparation and Identification All plasmid in Escherichia coli

Escherichia coli DH5 $\alpha$  In transformation amplification with the above mentioned Plasmid extraction kit extraction get. Preparation good plasmid Nano-drop 2000 Determination of concentration and purity, 1% Agarose gel electrophoresis identification in-Natural 20 Short-term save, -80 Long-term save.

1.3.2 Muscle in situ injection operation preparation the experimental group samples concentration 2 mg in mL<sup>-1</sup> Of PDNA Solution and concentration PEI Working Fluid by corresponding N/P Mixed which, DNA Total Quality MuG Incubated at room temperature Natural 20 min After formation N/P respectively 0.10 Of PEI/pDNA Complex. Then take different proportion of complex and 10  $\mu$ L 0.4% (W/v) Pluronic L64 Mixed with physiological saline dilution 40  $\mu$ L Pluronic L64 Final concentration 0.1%

(W/v) Mixture at room temperature static 5 min Formation L/P/D System (L/P/D-0.5, L/P/D-3, L/P/D-10) Which L/P/D-0.5 Group representative PEI And PDNA Of N/P = 0.5 L/P/D-3 Group N/P = 3 L/P/D-10 Group N/P = 10. To physiological saline group for negative control group, PDNA Group for positive control group 1 Pluronic L64/pDNA Mixture (L/D) Group for positive control group 2.

Mice 7/d Adaptation period after its on both sides of the Tibialis anterior muscle were was Hair Removal, Disinfection. With syringe respectively learn 40  $\mu$ L Control group solution and experimental Solution of the Group. Muscle injection along the parallel muscle fiber orientation needle about 2 mm<sup>2</sup>~5 S Complete Injection.

### 1.3.3 Report gene detection The 3A report gene detection system respectively from qualitative, Quantitative and gene expression evaluation of sustainability L/P/D System:

(1)  $\beta$ -Galactosidase qualitative detection: The organization to in situ staining of style the detection by observe the coloring range and color depth sentenced Broken gene expression situation. In accordance  $\beta$ -Galactosidase in situ Staining Kit instructions experimental including in ice in staining Stationary Liquid Solid

Set Natural 20 min With phosphate buffer (PBS) Washing 3 Times after fixed of sample the staining Processing 2 h More than finally with digital camera collection of Image.

(2) Fluorescein enzyme quantitative detection: The Luciferase Reporter Assay Kit Kit the detection. Specific steps including: Samples by ice uniform Pulp, -80 Night cracking and 4 12 000 r in min<sup>-1</sup> Centrifugal 3 min After take Natural 20  $\mu$ L Supernatant in don't light white 96 Hole ELISA plate in with multi-function microplate reader automatic and-like and read absorption value. At the same time BCA Protein Assay Kit The sample content determination homogenization Processing. Standard and sample with microplate reader Determination OD<sub>562</sub> Under the absorption value according to standard curve equation calculation the samples in protein content "with relative fluorescein enzyme activity (RLU/mg Protein) Said results.

3) Far-infrared fluorescence protein detection gene continuous expression situation: the living Imaging System of

muscle in situ injection after the first 7, 14 Days of red fluorescence protein E2-Crimson The expression of the detection. Detection before mice Tibialis anterior muscle hair removal processing with yellow light excitation in 600~

Nm The scanning fluorescence signal finally with instrument comes with software the fluorescence imaging analysis.

1.3.4 Organization pathology Detection The injection after the muscle organization and main organ (heart, Liver, Spleen, Lung, Renal) the tissue sections of and observe the evaluation cationic and Pluronic Combined with the gene transfer system of biological safety for clinical "with provide basis. Muscle injection 7/d After separation calf Tibialis anterior muscle or tail vein injection 4 d After anatomy out mice main organ PBS Washing after 4% (W/v) More pom Stationary Liquid in fixed 48 h After in ethanol and xylene solution in the gradient dehydration. Dehydration after of samples after paraffin embedding, Slice, Hematoxylin-Eosin (HE) Staining processing after with optical microscope.

1.3.5 L/P/D-0. 5 System in PEI/pDNA Complex of Particle Size, Potential and Morphology Characterization 3 MuG Or Natural 20 MuG pDNA And concentration

Ng in  $\cdot \text{mL}^{-1}$  PEI Working Fluid by corresponding N/P Mixed incubated at room temperature

Min After MilliQ Water dilution 1 mL. Will 1 mL Sample solution 1 cm Quartz Cuvette into Malvern nano-particle size and potential Analyzer (Zeta-Sizer Malvern) In by dynamic light scattering method

Dynamic light scattering (DLS) Okay N/P For 0. 5 An arcane PEI/pDNA Complex of Particle Size, Dispersion and surface potential the detection Samples DNA Final concentration 3 MuG In  $\text{mL}^{-1}$  Each sample repeat 3 Times.

Will complex adsorption in mica on and dry after "with atomic force microscope (AFM) Characterization its particle size and morphology characteristics Samples DNA Final concentration Natural 20 MuG In  $\text{mL}^{-1}$ .

## 1.4 Statistical Analysis

The Origin, Prism 5. 0 The data statistical analysis the group between single factors variance (One-way ANOVA) Or T Test Analysis,  $P < 0.05$  For difference have statistical significance.

## 2. Results

### 2.1 $\beta$ -Galactosidase qualitative and Fluorescein enzyme Quantitative Analysis

The is application cationic material construction efficient of skeletal muscle in situ Gene Transfer System of key. Muscle injection 7/d After in  $\beta$ -Galactosidase (Figure 1:A) And fluorescein Enzyme (Figure 1:B) Of expression level on, L/P/D-0. 5 The gene expression level of the group was significantly higher than that of the positive control group. 1. Positive control group 2. However, N/P (L/P/D-3 And L/P/D-10) Gene expression began to decline significantly. Numerically, L/P/D-0. 5 Group-mediated luciferase expression was positive in the control group. 1. 28. 6 Times, Positive control group 2. Of 2. 5 Times. However, when N/P Add 3. And 10 The expression of foreign genes was almost undetectable, and its value was reduced by at least 4. Order of magnitude. These results indicate that only at a specific low N/P Time, Pei To the system

Gene, inheritance, delivery and expression, yield, product, extreme shadow, sound. Therefore L/P/D-0. 5 For Optimization L/P/d System.

### 2.2 Expression of Fluorescent Protein Gene

Continuous Study of Exogenous Gene Expression in vivo is more convincing for assessing the intensity and persistence of gene expression. (Pu *et al.*, 2014). In vivo imaging, positive controls 1. And L/P/D-0. 5 Groups of Fluorescence signals can continue to express at least 2. Zhou Jian

Apparent Attenuation. L/P/D-0. 5 The fluorescence signal of the group was significantly higher than that of the positive control group. 1. (Figure 2.).

### 2.3 Histological Analysis

Intramuscular injection 7 d After, L/P/D-0. 5 Group and negative control group He No tissue lesions were found in the

stained sections (Figure 3.:A,B), Note less

Quantity PEI It is safe for skeletal muscle in situ Gene Transfer System. With N/P Increase, L/P/D-3 The group showed pathological changes such as muscle fiber degeneration and inflammatory cell infiltration (Figure 3.:C). When N/P Reach 10 Time, He Large infiltration of inflammatory cells in stained sections, Pathological Changes of lymphocyte infiltration and muscle fiber necrosis (Figure 3.:D).

Tail vein injection 4 d After that, the acute toxicity of the system was evaluated by observing the tissue sections of the main organs. L/P/D-0. 5 The results of the group were consistent with that of the negative control group (Figure 4.) That is consistent with the results of local injection.

/P/D-0. 5 It will not cause organ lesions, which further indicates that the system has good biocompatibility..

A. negative control group, B. L/P/D-0. 5 Group, C. L/P/D-3 Group, D. L/P/D-10 Group; The yellow arrow shows

2.4 L/P/D-0. 5 Characterization of complexes in the system [304 Nm  $\pm$  7. 6 nm, Multiple dispersion coefficient (Polydisperse) in- N/P For 0. 5 Of PEI/pDNA Complexes (PEI/pDNA-Dex, PDI) = 0. 43], And Zeta The potential is negative (-15. 9 eV  $\pm$  0. 5) The particle size is shown in Fig. 5.: A The proportional complex has nano-size 1. 4 EV) Show when N/P For 0. 5 Time, PEI With PDNA Formation

### 3. Discussion

Skeletal Muscle in situ Gene Transfer and Expression System for gene treatment provide the ideal of strategy the system of open allow will many kinds of methods combined with Application to get better of treatment effect. Non-ionic Style Material Pluronic Can improve the gene transfer efficiency main : (1) And Virus Carrier the to improve its safety and enhanced Gene Transfection Efficiency

Feldman *et al.* 1997; Dishart *et al.* 2003); (2) As an carrier modified Group (Jeon *et al.* 2003) Or auxiliary agent (Astafieva

Et al. 1996) Improve cationic non-Virus Vector in Serum/Physiological environment in stability and improve the gene transfection efficiency ; (3) Separate or and physical methods combined with can significantly enhance exogenous gene in muscle in situ Gene Transfer System in transfer/Expression Level (Gu, Rin 2000; Song *et al.* 2013; Liu *et al.* 2014). Pluronic May through many kinds of Molecular Mechanism promote exogenous gene transfer and expression such as by enhanced cell membrane permeability to enhance PDNA, Virus Carrier or cationic Material/DNA Complex of uptake (Gebhart *et al.* 2002); Promoting

PDNA In skeletal muscle in penetration, Distribution or into nuclear role and (Gu, Rin 2000; Pitard *et al.* 2002). In a kind Pluronic L64 Mediated-of efficient of skeletal muscle in gene transfer programme in, Pluronic L64 Use similar of phospholipid molecular structure and cell membrane each other role interference its integrity improve Membrane Permeability (Liu *et al.*

2014). However in the system in due to its Pluronic L64 Don't and PDNA Each other role not protection DNA In complex of in Environment in easy to be nuclease degradation difficult to smooth, Security transfer to target Cell in the transfer great to reduce the muscle in Gene Transfer Efficiency and serious of organization inflammation (Pitard *et al.* 2004; Burke & Pum 2008). It is suggested that with negative charge or electrically neutral of material/PDNA

Complex can avoid and ECM In electronegativity molecular of non-specific binding may be suitable for muscle in Gene Transfer System. In PEI And PDNA The composite system in by regulation cationic PEI And PDNA Of Proportion preparation the with different surface charge of Complex. So this research hope find a right PEI And PDNA Of proportion to formation overall with negative charge of complex to avoid and ECM In with negative charge molecular of each other role at the same time reduce PEI The amount of also can reduce even avoid local inflammation reaction and these all may be in favor of the objective gene to muscle cells in the transfer and Expression. This experimental results confirmed that only L/P/D-0. 5 The Group can significantly increase the expression level of foreign genes, and maintain the high level of expression within the monitoring time. N/P Increase, L/P/d The gene expression was dramatically reduced, even lower than the positive control group. 1.. At the same time, whether it is intramuscular injection or tail vein

injection, L/P/D-0. 5 They all show good biological compatibility.

It can be safely applied to skeletal muscle gene transfer to enhance the expression level of exogenous genes, and improve N/P Muscle tissue will have varying degrees of pathological changes, especially N/P Time (N/P = 10). The above-mentioned gene expression levels and biological safety data are consistent with the expected results, proving that in certain low-profile conditions, PEI/pDNA The electronegativity of the complex indeed helps to construct efficient and safe in situ Gene Transfer in skeletal muscle/Expression System. It is worth mentioning that this proportion is completely invalid in vitro gene transfection experiments, N/P Site (Schmidtwolf & Schmidtwolf, 2003). Meanwhile, In 10~15 Optimal transfection Condition. This also indicates that

Compared to nucleic acid molecules compressed into nanoscale, uncompressed Because the conditions required for delivery are very different, this may be related

PDNA Due to the loose molecular chain state, the efficiency of transmembrane Movement Different Environment. Will decrease (Godbey *et al.*, 1999).

A common nucleic acid protection strategy is to use cationic carriers to compress and protect negatively charged nucleic acid molecules through charge effect. As a recognized nucleic acid carrier, Branched Polyethylene imine is widely used in Gene Transfection in vitro, showing excellent transfection effect. (Xu *et al.*, 2009; He *et al.*, 2013). However, most in vivo experiments reveal the ineffective and even strong inhibitory effects of cationic materials on Gene Transfer and Expression in muscle. (TROs *et al.*, 2010; Song *et al.*, 2013; Pu *et al.*, 2014). This may be due to the difference between the in vitro cell culture environment and the complex in vivo muscle environment (RUponen *et al.*, 1999). Extracellular Matrix (Extracellular Matrix, ECM) A large number of negatively charged molecules can be combined with positively charged molecules. PEI/pDNA Complex, thereby interfering with the compound to the muscle

Because PEI Full compression DNA Time N/P For 3. (Dai *et al.*, 2011; Dai & Wu, 2012) So, N/P For 0. 5 When, in the compound DNA Can't be completely compressed, but a small amount PEI The addition DNA The structure of the nanocomposites is relatively close, forming a certain three-dimensional shape and negatively charged. Although 2. The particle size of the composites detected by different detection methods is different, and the error may be caused by different sample preparation conditions. AFM The images reflect the 3D structure of the dried nanoparticles, while DLS Nanoparticles need to be measured in solution state (Li *et al.*, 2001). We speculate that in L/P/D-0. 5 In the system, PEI/pDNA The electronegativity of the complex prevents it from being stranded ECM It is conducive to its transmission to the cells; 2)

Et al., 2015) Creating favorable conditions for complex molecules to enter cells; PEI Compression PDNA It forms a more compact complex molecule, which is more conducive to its passage through the permeability increased cell membrane and resist the degradation of nuclease. These conditions make L/P/D-0. 5 The transfer and expression efficiency of the system exceeded that of plasmid and Pluronic 164 The system of mediation has reached a new height.

In this study BALB/c Mice as models, validate an important concept: Will PDNA Compressed into negatively charged nano-composite particles, which can be applied and improved Pluronic 164 In situ Gene Transfer and Expression in skeletal muscle. How to promote the proof of this concept? DNA/Material complexes provide a solution to enter skeletal muscle cells, creating more efficient in-situ Gene Transfer/ The expression system will play a driving role in the design of a more appropriate material for molecular compression DNA To further improve the expression level of foreign genes, and may even push the system to the application level. In fact, in this study, the persistent high expression of red fluorescent protein in mouse skeletal muscle cells after one-time intramuscular injection has shown that the system has, Application Prospect of chronic diseases.

## References

1. Astafieva I, Maksimova I, Lukanidin E, Et al. 1996. Enhancement-ment of the polycation-mediated DNA uptake and cell transfection with Pluronic p85 block polymer[J]. FEBS letters, 389(3.):278-280.
2. Burke RS, Pun Sh. 2008. Extracellular barriers to in vivo PEI and pegated PEI polyplex-mediated gene delivery to the liver[J]. Bioconjugate chemistry, 19(3.):693-704.

3. Dai Z, Wu C. 2012. How does DNA complex polyethyleneimine with different chain lengths, topologies, aqueous solution mixtures? [J]. *Macromolecules* 45(10):4346-4353.
4. Feldman LJ, Pastore CJ, Aubailly N, et al. 1997. Improved efficiency of arterial gene transfer by use of poloxamer 407 as a vehicle. *Adenoviral Vectors* [J]. *Gene Therapy* 4(3):189-198.
5. Gebhart CL, Sriadibhatla S, Vinogradov S, et al. 2002. Design, formulation, polyplexes based Pluronic-polyethyleneimine conjugates. *Gene Transfer* [J]. *Bioconjugate Chemistry* 13(5):937-944.
6. Godbey WT, Wu K, Mikos AG. 1999. Size matters: Molecular weight affects efficiency of poly(ethyleneimine) as a gene delivery vehicle [J]. *Journal of Biomedical Materials Research* 45(3):268-275.
7. He Y, Cheng G, Xie L, et al. 2013. Polyethyleneimine/DNA polyplexes: reduction-sensitive hyaluronic acid derivatives shielding targeted gene delivery [J]. *Biomaterials* 34(4):1235-1245.
8. Lauritzen HP, Reynet C, Schjerling P, et al. 2002. Gene gun bombardment-mediated expression and location of EGFP-tagged GLUT4 in skeleton muscle fibers in vivo [J]. *Pflügers Archiv* 444(6):710-721.
9. Lee JI, Lo CW, Ka SM, et al. 2012. Prolonging the expression duration of ultrasound-mediated gene transfer using PEG nanoparticles [J]. *Journal of Controlled Release* 160(1):64-71.
10. Liu S, Ma L, Tan R, et al. 2014. Safe and efficient local gene delivery into skeleton muscle via a combination of Pluronic 164 and modified electrotransfer [J]. *Gene Therapy* 21(6):558-565.
11. Lu QL, Bougharios G, Partridge TA. 2003a. Non-viral gene delivery in skeletal muscle: A protein factory [J]. *Gene Therapy* 10(2):131-142.
12. MIR LMB, Bureau MF, Rangara R, et al. 1998. Long-term, high level in vivo gene expression after electric pulse-mediated gene transfer into skeleton muscle [J]. *Comptes Rendus de l'Académie des Sciences Série Sciences de la Vie* 321(11):893-899.
13. Pitard B, Belloroufa M, Lambert O, et al. 2004. Negatively charged self-binding DNA/poloxamine nanospheres for in vivo gene transfer [J]. *Nucleus Kids Research* 32(20):E159. Doi: 10.1093/NAR/gnh153.
14. Pitard B, Pollard H, Agbulut O, et al. 2002. [J]. *Human Gene Therapy* 13(14):1767-1775.
15. Pu L, Gene Y, Liu S, et al. 2014. Electroneutralized am-philic triblock copolymer with a peptide dendron for efficient muscular gene delivery [J]. *ACS Applied Materials & Interfaces* 6(17):15344-15351.
16. Schmidwolf G, Schmidwolf IG. 2003. Non-viral, hybrid vectors. *Human Gene Therapy: An Update* [J]. *Trends in Molecular Medicine* 9(2):67-72.