

Nile Tilapia Xbp1-S Gene Cloning and Expression Analysis

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Abstract: To learn about Nile tilapia (*Oreochromis niloticus*) Cell Transcription Factor Xbp1-S (OnXbp1-S) Sequence characteristics of the Gene and Its Role in *Streptococcus agalactiae* (*Streptomyces alactolyticus*) Stress and in B Role in cell differentiation, Application Race Obtained by cloning technology OnXbp1-S Full Length of Gene 1380 BP Including open reading frame ORF For 1155 BP, 5' Wang Yi Terminal non-coding region (5') Wang Yi UTR

The organization in have expression, MRNA Level on in liver in expression highest protein level in thymus in expression highest and in muscle in expression minimum. No milk *Streptococcus* stress after, OnXbp1-S Gene in liver and spleen in expression trend similar were in stress during appear expression was up-regulated in 192 h The peak. In addition immune was analysis found, OnXbp1-S Factor in different differentiation degree B Cells subclass in Expression Difference in mature B Cells in present high expression and in not mature B Cells in almost don't expression. Research results show that, OnXbp1-S Participate in Nile tilapia on no milk streptococcus of immune defense and in B Cells Differentiation role. This study will for further study OnXbp1-S Factor response pathogenic bacteria infection of mechanism and promote B Cells Differentiation Mechanism provide theory basis.

Keywords: Nile tilapia; Xbp1-S; Gene cloning; Organization expression; no milk streptococcus

Xbp1-S is a highly conservative protein from low yeast to Higher Nursing animal in there (Liou et al 1990). In the bony fish, Xbp1-S The Research of fish, including zebrafish, is gradually increasing. (Dario rerio) (Hu et al, 2007; Bennett et al, 2007) Carp (*Cyprinus carpio*) (Peng shaoqing and others, 2013) Takifugu with red fin (*Takifugu rubripes*) (Ohtani) et al, 2006) Rainbow trout (*Oncorhynchus mykiss*) (Barr et al, 2011) And Atlantic salmon (*Salmo salar* Leong et al, 2010) Etc. Hu Wait. (2007) In zebrafish embryonic cells, when UPR When dealing with endoplasmic reticulum stress, XBP1 Can pass IGF1/Akt Activation of apoptosis signal plays an inhibitory role; Barr Wait. (2011) Utilization XBP1 Identification of different degree of differentiation as a marker B Cell subclass; Zollo Wait. (2005, 2008) Utilization XBP1 And Pax-5 And other Transcription Factors on the kidney of rainbow trout, and further reveal B Development and differentiation of cells XBP1 Expression is related. But about Nile tilapia Xbp1-S Research has not been reported. In addition, compared with mammals

Xbp1-S The role of the host in response to pathogen infection remains to be further clarified.

Tilapia is one of the high-quality aquaculture varieties recommended by the United Nations. China is the largest tilapia producer. In recent years, large-scale breeding, high-density breeding, water pollution and so on have led to a large outbreak of tilapia fish disease, such as streptococcus, causing huge economic losses.

(Rumai new, 2010). In this study, Nile tilapia was selected as the experimental fish. Xbp1-S Gene cDNA Gene and

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protein levels were used to analyze the spatio-temporal expression patterns of tilapia tissues after *Streptococcus agalactiae* stress. The expression level of cell subclasses in Nile tilapia Xbp1-S Role and regulation in the prevention of streptococcus infection To provide a theoretical basis for the mechanism of cell development and differentiation.

1. Materials and Methods

1.1 Experimental materials

Experimental fish were purchased from Guangdong fishery germplasm protection base, healthy Nile

The weight of tilapia is (200 ± 10) g, Body length (13 ± 2) cm. Breeding Temperature is (29 ± 1) °C. Feed, air pump oxygen supply, feeding 14 d Make it fully adapt to the breeding environment, observe the fish body appearance and activity ability, and confirm that it is completely healthy for the experiment. *Streptococcus agalactiae* strain as this experiment

Room preservation strains. PCR Reagents for reaction systems, PMD-18T Vector, Restriction enzymes, Trizol Reverse Transcription Kit (Primescript™ RT agent kit with gDNA eraser) And Fluorescence Quantitative Kit (SYBR premix exTaq™) Both purchased from Takara Company; *E. coli* DH5 α Receptive, *E. coli* BL21 (DE3) Receptive, Gel recovery kit and plasmid extraction kit (Tianprep mini plasmid kit) Purchased from Tiangen Company; PET32a () vector Preserved by our laboratory, Smart™ RACE cDNA Amplification Kit Purchased from BD Biosciences clontech Company. Trizol, 2. Wang Yi Prime mix, ExTaq, 10 Wang Yi Buffer, DNTP, PMD 18-T Vector, Solution I And reverse transcription kit purchase

Takara Company. Ampicillin, chloroform, isopropanol and other reagents were purchased from Guangzhou Chemical Reagent Company, and the rest of the experimental reagents were made in China. Primer (Table 1) It was synthesized by Huada Gene Technology Service Co., Ltd.

1.2 Experimental Method

1.2.1 General RNA Extraction and cDNA Synthesis of the first chain Adopted Trizol (Invitrogen) Law General RNA Extraction. Take the healthy Nile tilapia head kidney, fully ground with a homogenizer, and follow Trizol Reagent manual extraction total RNA, RNA Integrity and purity 1% Agarose gel electrophoresis and micro Spectrophotometer Nano2000 Detect its concentration, Wang Yi 80C. Preservation. Reverse Transcription synthesis using a common reverse transcription Kit cDNA Template, using Smart™ RACE cDNA

Amplification kit Will total RNA Reverse Transcription 5. Wang Yi, 3. Wang Yi-RACE cDNA Template (Yao Et al, 2016). After reverse transcription, the validity of the template was tested Beta-Actin As an internal reference Gene PCR After Qualification-20C. Preservation.

1.2.2 Nile tilapia Xbp1-S Full Length of Gene cDNA Cloning

AND SEQUENCING In NCBI Search for Nile tilapia XBP1 Genes, based on the predicted Sequence (Xm_013276886.1), Application Software Primer Premier 5.0 Carry Primer Xbp1-F1 And Xbp1-R1 Design (Table 1) To Nile tilapia head kidney cDNA As a template Xbp1-F1 And Xbp1-R1 For primer Xbp1-S Of cDNA The amplification, PCR The reaction system is 25 μ L, Reaction procedure: 94 Pre-denaturation 3 min; 94 Denaturation 30 s; 58.0 Annealing 30 s; 72C Extension 1.0 min;

A loop. Amplified PCR Product Sutra 1% After agarose gel electrophoresis, it was sent to Beijing Liuhe Hua big Gene company for sequencing. Sequencing

Results NCBI (<http://www.ncbi.nlm.nih.gov>) Compare and get Nile tilapia Xbp1-S Of cDNA Sequence.

1.2.3 Nile tilapia Xbp1-S Gene 3. Wang Yi And 5. Wang Yi-Race Amplification

According to the obtained Xbp1-S Of cDNA Sequence, design Xbp1-S

Of 5. Wang Yi-RACE PCR And 3. Wang Yi-RACE PCR Primer Xbp1-F2, Xbp1-F3, Xbp1-R2 And The Xbp1-R3 (Table 1), Using nested PCR

Method proceed 3. Wang Yi And 5. Wang Yi-Race The amplification Xbp1-F2, Xbp1-R2 And UP Mixed Primer (Mixed concentration Reference Kit Manual) First-round amplification and then first-round Amplification PCR Product

Dilution 10 times as the second round, dilute, plate, and then use Xbp1-F3, Xbp1-R3. The second round of amplification was carried out with primers. 20 μ L PCR Reaction System (Yao et al, 2016). Reaction Procedure:

Pre-denaturation 3 min; 94 Denaturation 30 s; 68 Annealing 30 s;

C Extension 1 min, 35 A loop. Amplified PCR Product. After agarose gel electrophoresis, it was sent to Beijing Liuhe Hua big Gene company for sequencing. Sequencing results and Xbp1-S of cDNA Sequence alignment results are shown in Fig. 1., Find the start password and PolyA Add tail signal.

1.2.4 Nile tilapia Xbp1-S Sequence analysis With DNASTAR

In software Seqman Program to clone the carrier, Will 3. Wang Yi, 5. Wang Yi. Combination of sequencing results. cDNA Sequence splicing analysis, further confirmation. Xbp1-S Amplification obtained cDNA The full length. With NCBI Website Blast Sequence Homology analysis of the encoded protein; Bioedit Soft

Of Nile tilapia and other fish Xbp1-S Multiple alignment of the amino acid sequence; Application Mega 6.0 Software, with ortho-junction Method (NJ Law) Build a phylogenetic tree and Bootstrap Repeat 1000 Calculate the confidence of each branch. Prediction of protein Physicochemical Properties ProtParam Software (<http://web.expasy.org/protparam>), So that

NCBI Site conservative domain (CDD) Database (<http://www.ncbi.nlm.nih.gov/structure/cdd/wrpsb.cgi>) Prediction of conserved domains for amino and basic acid sequences.

1.2.5 Nile tilapia Xbp1-S Preparation of polyclonal antibody According

Xbp1-S of cDNA Sequence of the zinc finger structure. 504 BPP Prokaryotic Expression Analysis and construction PET32A-XBP1 Protein Expression Vector and import BL21 Specific Expression of about 40 kDa Destination fragment (Contains Ppet32a Molecular Weight of Carrier). Then, the purified recombinant protein was concentrated with the Freund's adjuvant. Sigma Mixed, intraperitoneal immune mice (BALB/c, 8 Zhou), 2. Zhou immunization 1. Continuous immunization 4. For the first time, Freund's complete adjuvant was used, and the Freund's incomplete adjuvant was used again. 50 μ g/Only. After getting the antiserum, the resistance was detected. Elisa Check its titer. In the first 4. After immunization 4 D (Highest antibody titer) The antiserum was obtained from the tail vein for protein detection and immunohistochemical staining.

1.2.6 Nile tilapia Xbp1-S Organization expression Fluorescence Quantitative PCR Method detection XBP1 In MRNA Expression Level in healthy tilapia tissues, using Western-blot Method detection XBP1 Protein distribution in different tissues. Take healthy 3. The head kidney, posterior kidney, spleen, skin, Gill, intestine, thymus, liver and muscle tissue of tilapia tail Nile-80C. Preservation. And organizations RNA And tissue protein extraction. RT-PCR

Western-blot Detection of Tissue proteins. For RT-PCR Primers for detecting the expression in different tissues of Nile tilapia 1.. Reaction Conditions:

95C 3 min; 95C 1 min; 60C 30 s; 72C 30 s, 35 A loop. Repeat for each sample 3. Time, according C_T (Threshold cycles)

Value $(15) < C_T < 35$) To Beta-Actin Gene as internal reference, using formula 2^{-C_T} Draw each sample XBP1 Relative expression of genes.

1.2.7 Streptococcus agalactius stress test Take 60 Niro Nakai

Tilapia randomly divided 2. Stress group and control group. Intraperitoneal injection in stress group 100 μ L Concentration is 5×10^5 Cfu/ μ L Streptococcus agalactiae (Tellez-Bañuelos et al, 2009; Gan et al, 2015) Intraperitoneal injection in control group 100 μ L Sterile PBS, Injection 0, 6, 24, 48, 192, 264 h. After treatment, the stress group and the control group were randomly selected 3. The tail fish is frozen in liquid nitrogen and preserved in -80 Refrigerator for subsequent organizations XBP1 Gene's RT-PCR Analysis and detection. RT-PCR All rights reserved. <http://www.cnki.net>

1.2.8 Xbp1 Immune was Select Health Nile tilapia of head kidney white blood cells for immune was experimental. Head renal white blood cells of Separation Using Percoll Density gradient separation the according to Nile tilapia B Cells of proportion respectively preparation Percoll 40, 50, 60 And 70 Reagent superposition after Percoll Density gradient separation liquid to separation development of differentiation degree different B Cells subclass. In super-clean workbench take health Nile tilapia of head kidney and join 3 ml 1640 Cell culture medium the full grinding learn supernatant removal big of organization block spare. According to "with manual first Percoll (GE

Healthcare)Liquid10 PBS (Na_2PO_4 In 2 H_2O 2.286g Na_2HPO_4 In 12 H_2O 29.14g NaCl 87.75g) 9:1Dilution for sparePercollStorage liquid then1 PBSFurtherPercoll 40,50,60And70The superposition each layer SeparationBCells. The separation of cells further cells smear afterEnVisionTMImmune organization chemical two-step main steps including: slice with two toluene dewaxing gradient ethanol off benzene to water hydration;3%Hydrogen peroxide solution at room temperature to avoid light Incubation25 minBlocking endogenous peroxidase;3% BSAAt room temperature closed30 minAfter drop Add a,4 °C Incubation night; fluorescence two anti-incubated at room temperature30 min; Fluorescence microscope microscopic examination positive cells significantly of fluorescence and the image collection of analysis.

2. Results and Analysis

2.1 OnXbp1-SGeneCDNAFull-length Cloning and Sequence Analysis

According to acquiredORFSequence further design specificRACEPrimersXbp1-F2,Xbp1-F3AndXbp1-R2AndXbp1-R3 (Table1).Xbp1-F2AndXbp1-R2And general primersUPMAndXbp1-F3AndXbp1-R3And general primersNUPRespectively pair

3 -RaceAnd5 -RaceAmplification(YaoEt al2016)Of products were confirmed by sequencing Panel after get Nile tilapia gene full-lengthCDNASequence

1155 BP3 End non-coding region(3 'UTR)Long98 bp5 End non-coding region(5 UTR)Long127 bp3 ContainsPolyATail and more poly adenylateAataaAnd tail Signal(Figure1). Amino acid sequence analysis shows that,OnXbp1-SGene can Coding384A amino acid residues the derivation of molecular weight41.32 kDaTheory isoelectric point4.36. By protein of function forecast indicates that,OnXbp1-SContains174 bpOf zinc refers to structure regional(Figure1Underline part)The regional is compare the conservative protein structure the protein belongs to alkaline leucine zip structure protein.

2.2 OnXbp1-SGene of homology analysis

UseBioeditSoftware of Nile tilapiaXbp1Gene encoding the amino acid sequence and red takifugu, rainbow trout, and the European food with Mu Li(*Ostrea edulis*), Spot zebrafish, spots bird eel(*Lepisosteus*

Oculatus), Mexico fat carp(*Astyanax mexicanus*), Green (*Oryzias latipes*)(*Oryzias latipes*), Antarctic cod(*Notothenia coriiceps*), Jungle Fowl

(*Gallus gallus*), Mice(*Mus musculus*)And people(*Homo sapiens*)The homologous comparison and different species in zinc refers to the structure of homology up79% (Figure2). As shown in figure2Shown in black shadow said homology greater75%Of Regional Main is concentrated in different species of zinc refers to the structure of conservative regional. UseMEGA 6.0Software to have published of lactation animal and common of fish the system evolution analysis display Nile tilapiaXbp1And Antarctic cod belong to a branch homology highest and other invertebratesXbp1Together for a class. But there are obvious differentiation in mammals, amphibians, birds and fishes.(Figure3).

2.3 Expression and Purification of prokaryotic protein and preparation of Its Polyclonal Antibody

Nile tilapiaOnXbp1-SGene Fragment as template, using specific primersXbp1-F4,R4 (Table1)Successfully amplified to length

BPTo connect the destination fragmentPET32aPlasmid, obtain the recombinant plasmidPET32a-Xbp1. Transfer recombinant plasmid into *Escherichia coli*BL21 (DE3)After induction, the positive clones were sequenced correctly and the expression of the target protein before induction was weak.(Figure4)The size of the bands that were expressed after induction was about40 kDaConsistent with the predicted molecular weight of the fusion protein(Including target protein18 kDaAndPET-32aTag Protein21 kDa). The results of Soluble analysis showed that the fusion protein mainly existed in the form of inclusion bodies. JingSDS-PAGEDetection, purification of proteins in40 kDaThere is a

single band. With ResistanceHis-tagMonoclonal Antibodies AgainstWestern-blotThe purified protein was detected in
KDaThere is a single band atOnXbp1-SExpressed fusion protein. The purified protein was concentrated, and the
mice were immunized with antigen and antiserum was obtained.ElisaThe antibody titer was800,000 (units/ml)Shown
that the prepared mice were resistant to tilapia

Xbp1-SPolyclonal antiserum had good effect.

2.4 Nile tilapiaXbp1-STissue Distribution of gene and protein levels and stress expression of Streptococcus

AdoptedReal-time qpcrMethod detectionOnXbp1-SMRNAExpression in different tissues of healthy Nile tilapia;
PreparedOnXbp1-SPimaWestern-blotDetectionOnxpb1

Protein Expression levels in different tissues.MRNAThe results of relative expression showed that,OnXbp1-SIn the
Detected8.The expression in tissues, from low to high, followed by muscle<Thymus<Spleen<Skin<Gill<Chang<Head
kidney<The highest expression was found in the liver and in the head kidney.

After intraperitoneal injection of streptococcus, compared with the control group, the liver and spleen were
up-regulated at different time points, and the trend was basically the same. Early intraperitoneal injection6
hAfter,OnXbp1-SGene expression was significantly increased, and24 h

48 hThe expression192 h.To the highest level of Gene Expression(Figure6a, Figure6b)And then in264 h.Gradually
dropped down and tended to the level of the control group.

2.5 Nile tilapiaXbp1-SAt differentBIimmunohistochemical analysis of cell subtypes

AccordingZwolloWait.(2005)Reported.PercollTilapia head kidney was isolated by density separation
method.BCells:Percoll 40Cell(Plasma cells, fig.7A),Percoll 50Cell(More than puree

Cell, graph7B),Percoll 60Cell(Initial differentiation matureBCell, graph7c), AndPercoll 70Cell(Mostly
immatureBCell, graph7D). As shown in Fig.7.As shown,OnXbp1-SProtein in differentBThe expression level of the cell
subclass can be shown by its red fluorescence.

Change from low to high isPercoll 70 <Percoll 60 <Percoll 50 <Percoll 40OnXbp1-SThe highest expression in
plasma cells, the highest level of expression in primary plasma cellsBCells have a little expression, but in
immatureBCells are barely expressed.

3. Discussion

Cell Transcription FactorXBP1Expression andBThe development and differentiation of cells and the expression
and secretion of antibodies are closely related.(ZwolloEt al, 2005,2008; TELLIEREt al, 2016).XBP1It is one of the
major mediators of protein folding ability in endoplasmic reticulum stress.2.Different types of
cuts(Xbp1-SAndXbp1-U)Where,Xbp1-SHighly transcribed(ZhouEt al, 2008). Endoplasmic Reticulum regulates protein
folding, post-transcriptional modification gene expression and other functions,If a wrong folding occurs, it will lead to
an anti-apoptotic reaction and promote the unlimited proliferation of tumor cells.(Hendershot, 2004; RobinsonEt al,
1994). Therefore,XBP1ResearchBThe development, differentiation and regulation of Cells and Their Immune Function
in the defense of pathogenic bacteria infection are of great significance.

This study cloned Nile tilapiaOnXbp1-SAnd the prokaryotic expression system of exogenous protein was
used.(SalinasEt al, 2011)Successful buildOnXbp1-SIn Vitro Expression System, polyclonal antibody was
prepared,OnXbp1-SIn normal tissues of tilapia niloticus; in the stress period of Streptococcus,OnXbp1-SThe expression
in liver and spleen was up-regulated and the reaction tended to be synchronous.OnXbp1-SFor plasma cells that are
matureBRegulation of cell development and differentiation.

XBP1It is widely expressed in mammalian tissues, especially in hepatocytes.(LiouEt al, 1990; calfonEt al, 2002).
The results showed that Nile tilapiaOnXbp1-SIt is expressed in various detection organizations.MRNA

The level is mainly expressed in the liver, which is similar to the discovery in mammals. However, high protein

expression on the protein level is mainly in the thymus and head kidney, suggesting that Xbp1-EXPRESSION AND IMMUNE CELLS(B)Cell and TCell) There may be some correlation (ZwolloEt al, 2005,2008; TELLIEREt al, 2016). Development and differentiation of the Main Immune Cells in the immune system Xbp1-S The positive correlation of expression reflects Xbp1-S Transcription factors may play an important role in the immune response, because Xbp1-S Cytokines are not only involved in humoral immunity, but also may be involved in the promotion of cellular immunity. Streptococcus agalactius stress test showed that after stress 264 h. During, On Xbp1-S It was up-regulated in spleen and liver. Intraperitoneal injection of Streptococcus 6 h After the liver and spleen 4) On Xbp1-S mRNA The first peak was a significant increase in expression, which indicated that this factor might be involved in the natural immune response. The second more significant expression peak occurred after Streptococcus stress. 192 h (8 d), This significant increase is likely due to immune cells involved in the adaptive immune response (B)Cell and TCell) China Xbp1-S The expression of factor was significantly up-regulated.

The head kidney of the bony fish is similar to the bone marrow of mammals, having hematopoietic function and is a site for the development of immune cells. Therefore, the head kidney tissue contains immature blood cells of different blood lines. (FengeEt al, 1986; ZapataEt al, 1995). The head kidney is an important place for the occurrence, development, differentiation and proliferation of immune cells.

Organ (KaattariEt al, 1985; bromageEt al, 2004; yeEt al, 2010; MAEt al, 2013). In this study, the head kidney tissue of Nile tilapia was Percoll Density gradient separation method to obtain different developmental Differentiation B Cell subclass (ZwolloEt al, 2005) Immunohistochemical staining was used to detect the fluorescence intensity of cells. On Xbp1-S Highly expressed in mature plasma cells and protoplasmic cells with high degree of differentiation. Naïve B Cells and immature B The expression of cells was low. Nile tilapia

Cell subclasses expressed at different levels On Xbp1-S Phenomenon, with the reported red fin takifugu (OhtaniEt al, 2006) And rainbow trout (ZwolloEt al, 2008) Similar. B Cell Differentiation and Xbp1-S The positive correlation of factor expression indicates that this factor plays an important role in promoting the development of mature plasma cells and B Cell Differentiation plays a positive role in regulating the differentiation of protoplasmic cells. (YeEt al, 2011). Xbp1-S Immune Function especially B The role of cell differentiation is conducive to the exploration of humoral immunity B Regulation Mechanism of cell development and differentiation. In conclusion, this study will provide a basis for further study of Nile tilapia. Xbp1-S Mechanism and promotion of factor response to pathogen infection B Provide a theoretical basis for cell differentiation mechanism.

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